

escape interference line can be seen at $1332-511=821$, and the double escape interference line at $1332-1022=310$.

- 4.4 **Summation Interference:** When high gamma emission rates are seen, sample summation can occur. Prominent in geometries close to detection and in low energy range (i.e., 10,000 cps at 88 KeV, 15,000 cps at 210 KeV), a summation interference can be seen at $88+88=176$ KeV, $210+210=420$ KeV, $210+88=298$ KeV.
- 4.5 **False Positive:** An isotope that has failed one or more of several tests including half-life, abundance, and energy tolerance (± 2 KeV)
- 4.6 **Abundance Test:** The test where the software calculates the total possible lines from the library and checks to see how many were actually seen. The cutoff for a positive identification is 75%.
- 4.7 **Energy Tolerance:** The test where the software checks the energy line in the spectrum to see if it is within the energy tolerance setting. (The standard setting is 2 KeV.) If it is within this setting then the line is associated with that nuclide. The energy line can be associated with more than one nuclide.
- 4.8 **Half-Life Test:** The test to determine if the half-life of the isotope is long enough not to have decayed away. The half-life of the sample is the time from sample date to analysis date plus 1/2 the count time. A limit of no more than eight half-life is the standard setting.
- 4.9 **Key Line:** The line chosen by the builder of the library to be the prominent line of the isotope. This line is used in the MDA table for purposes of calculating activity, error and MDA. For non-identified isotopes the key line is used as the basis for calculating a region around the key line and then calculating activity error and MDA. Usually this line is the most abundant line on a line that is relatively free from interference.
- 4.10 **Abundance:** The branching ratio or ratio of disintegration of the isotope at a particular energy. For example, Cobalt-60 has an abundance, or branching ratio, of 99% at 1332 KeV.
- 4.11 **Accuracy:** The error of the reported result due to the counting statistics of the instrument used for quantification.
- 4.12 **Back Scatter:** The detection of a count that occurs when an event interacts with counting materials, changes direction, and scatters back to the detector.

5.0 METHOD VARIATIONS

Modifications to the procedure are limited to GEL's use of additional isotopes for the daily calibration check and the inclusion of a more stringent calibration and resolution periodicity.

6.0 SAFETY PRECAUTIONS AND WARNINGS

- 6.1 Keep hands free from moving parts of canning device and Gamma shields.
- 6.2 Personnel performing this analytical procedure are trained in and follow the safe laboratory practices outlined in the Safety, Health and Chemical Hygiene Plan, GL-LB-N-001.
- 6.3 Personnel handling radioactive materials are trained in and follow the procedures outlined in GL-RAD-S-004 for Radioactive Material Handling.

10.1.2 Fill the appropriate container with sample prepared from step 10.1.1 using the following steps as a guideline:

10.1.2.1 If Ra-226 analysis is required, the sample is placed in a 100cc can for in-growth.

NOTE: It is recommended that in-growth be allowed 14 days to quantify Ra-226. Shorter intervals can be used at the request of the client. However, shorter in-growth periods may decrease the accuracy of the data. If there is insufficient mass of sample to fill the 100cc can, contact the team or group leader.

10.1.2.2 All homogenized samples shall be placed in the 100cc can. Determine the net weight of the sample. If the net weight is less than 55 grams or greater than 190 grams, contact the team or group leader to determine the appropriate counting container. Record sample weight and date on sample container.

10.1.2.3 If there is insufficient sample to fill the 100cc can, place sample in the 10cc petri dish, cap and seal. Record sample weight and date on sample container.

10.1.2.4 If there is insufficient sample to fill the 10cc petri dish, perform the following digestion process:

10.1.2.4.1 Weigh out an appropriate aliquot into a labeled teflon beaker. Record this weight on the sample container.

10.1.2.4.2 Add 10 mL of concentrated nitric acid to each sample.

10.1.2.4.3 Place samples on medium heat (~300 °F) and cover each sample with a teflon lid. Reflux all samples for 30 minutes.

10.1.2.4.4 Remove teflon lids and add 5 mL concentrated hydrochloric acid and 10 mL hydrofluoric acid to each sample. Cover samples and reflux for 120 minutes.

10.1.2.4.5 Remove teflon lids and allow samples to evaporate to dryness.

10.1.2.4.6 Add 5 mL of concentrated nitric acid and evaporate to dryness.

10.1.2.4.7 Repeat Step 10.3.6.

10.1.2.4.8 Add 5 mL of concentrated nitric acid to the dry samples. Place the samples back on the hotplate long enough so that the dried sample dissolves into the acid.

10.1.2.4.9 Transfer solution to a 500 mL vessel and dilute to 500 mL. Record original sample mass and diluted volume on sample

10.2 Water sample preparation

$$\text{Decay} = \left(\frac{T_{1/2\text{err}}}{T_{1/2}} \right)^2 * \left[\frac{\lambda E_r}{1 - e^{-\lambda E_r}} - \lambda (T_s + E_r) - 1 \right]$$

- 15.4 The method MDA in pCi/g or pCi/L are calculated according to the following equations:

$$\text{MDA (pCi/unit)} = \frac{d * (2.71 + 4.66 \sqrt{\text{cpm}_b * \text{ct}})}{2.22 * E * V * B * \text{ct}}$$

Where:

- A = net peak area (counts)
- ABS = relative absorption factor
- B = abundance (gammas/disintegration)
- E = counting Efficiency (counts/gamma)
- V = sample volume (grams or liters)
- ct = sample count time (minutes)
- d = decay factor = $d = \frac{1}{e^{-\lambda}}$

- 15.5 The absorption factor is calculated by the following equations:

$$I_1 = \frac{\ln((SS\text{cpm} - S\text{cpm})/EC\text{cpm})}{(((SS\text{cpm} - S\text{cpm})/EC\text{cpm}) - 1)}$$

$$I_0 = \frac{\ln((SST\text{cpm} - ST\text{cpm})/EC\text{cpm})}{(((SST\text{cpm} - S\text{cpm})/EC\text{cpm}) - 1)}$$

$$\text{ABS} = \frac{I_1}{I_0}$$

Where:

- SScpm = sample plus the source cpm at the region of interest
- Scpm = sample cpm at the region of interest
- ECcpm = source cpm on the empty can at the region of interest
- ln = natural logarithm
- SSTcpm = standard plus the source cpm at the region of interest
- Stcpm = standard cpm at the region of interest

- 15.6 The VAX operating system will report the following information with each completed sample:

- 15.6.1 The nuclide identification report
- 15.6.2 The minimum detectable activity report
- 15.6.3 The peak search report.

- 15.7 The following criteria are used to accept a reported gamma isotope from the NID report:

- 15.7.1 The peak FWHM should be less than 2 KeV.

Appendix E

Analytical Results

